

Editorial Focus

Cell-derived microparticles: A mediator of inflammation in aortic valve stenosis?

Yeon S. Ahn, Wenche Jy, Laurence L. Horstman, Joaquin J. Jimenez

Wallace Coulter Platelet Laboratory, Miller School of Medicine, University of Miami, Miami, Florida, USA

Aortic valve stenosis (AVS) is quite common, affecting some 30% of those over age 65 (1). It frequently progresses to severe obstruction necessitating surgical repair or replacement of the stenotic valve. Annually more than 50,000 aortic valves are replaced surgically in the USA, posing common and serious medical problems (2), yet our knowledge of the pathogenesis and optimal treatment of this condition is surprisingly limited. It may represent a manifestation of atherosclerosis affecting the endothelium lining the heart valve but this is still debated. There is mounting evidence that AVS is associated with systemic inflammation (3, 4). Some but not all studies found an association of AVS with elevated C-reactive protein (CRP) (3) and active atheroinflammatory processes (4), implying a role of inflammation in the pathogenesis of AVS.

Even if true, however, there is little insight on exactly how or why mechanical perturbation of blood flow due to AVS leads to systemic inflammation, or what inflammatory processes promote the initiation and progression of AVS. Elucidation of this key issue could provide a rational basis for prevention and therapeutic strategies.

The intriguing report by Diehl et al. in this issue of *Thrombosis and Haemostasis* (5) may offer an insight into this problem, and may be of potential clinical importance. They hypothesize that AVS induces generation of circulating cell-derived microparticles (MP) by shear stress, and that the resulting MP then promote systemic inflammation. Briefly, their scenario goes like this: high shear stress generates platelet microparticles (PMP), which in turn interact with leukocytes to activate them and elicit release of leukocyte MP (LMP) and monocyte MP (MoMP). These procoagulant and inflammatory MP, along with activated monocytes, can induce endothelial injury at the lining of the heart valve, thereby setting up a “vicious cycle,” as the authors put it, leading to further calcific stenosis of aortic valves as seen in AVS. Endothelial cell injury is reflected by release of endothelial MP (EMP) (6).

This hypothesis is novel and intuitively appealing. To support it, they measured MP species including PMP, LMP, and EMP in 22 AVS patients, and report a significant increase in all of them relative to controls. In parallel, elevation of markers of systemic

inflammation was observed, such as soluble P-selectin, interleukin-6, and activation of monocytes. These data convincingly support the hypothesis, although falling short of conclusively establishing a cause-effect relationship between MP-cell interaction and progression of AVS.

In addition, since this is a cross-sectional study, more definitive support for an active role of MP in the pathogenesis of AVS will require a prospective longitudinal study correlating MP-cell interaction with progression of AVS, as the authors acknowledge in their discussion.

Thus, a central theme of this paper is the physiological relevance of MP-cell interactions. They demonstrate copious production of PMP in AVS, and correlation of PMP levels with valvular shear stress *in vivo*. Others have previously shown *in vitro* that PMP are readily generated by abnormally high shearing forces (7–9). It has also been shown that PMP can induce leukocyte activation (10, 11), and can act directly on endothelial cells (8, 12). High shear can also induce endothelial activation and release of EMP (13); and in turn, EMP can activate leukocytes (14–17). Indeed, EMP were reported to act on the endothelial cells that produce them (18). Mixed MP isolated from patients were shown to be deleterious on cells *in vitro* or *ex vivo* (19, 20). Finally, the LMP elicited by PMP or other MP are known to be injurious to endothelial cells (21, 22).

In view of these and other reports cited by Diehl et al. (5), there is ample evidence *in vitro* to support their hypothesis, that interactions of shear-generated MP with blood cells and endothelium could be pivotal in the progression of AVS.

Their report struck a chord with us because of our anecdotal observation some years back of a patient with severe AVS, who refused surgical intervention. Her PMP counts were among the highest we had ever seen. She suffered from numerous recurrent transient ischemic attacks, progressing to advanced dementia. Thus, it is gratifying to see our observation confirmed in a significant number of patients by the work by Diehl et al.

What implications might this report have for therapeutic strategies? If the hypothesis of Diehl et al. reported in this issue is correct, then it may be time to begin thinking about ways to modulate MP-mediated injury. Two approaches come to mind,

Correspondence to:

Yeon S. Ahn
Wallace Coulter Platelet Laboratory
1475 NW 12 Ave
Miami, FL 33136, USA
Tel.: +1 305 243 6606, Fax: +1 305 243 4975
E-mail: yahn@med.miami.edu

Received March 6, 2008
Accepted March 6, 2008

Prepublished online March 12, 2008
doi:10.1160/TH08-03-0135

Thromb Haemost 2008; 99: 657–658

first being to inhibit their interaction with cells, the second being to inhibit MP release. Regarding the first, several strategies are now being tried or considered which block specific cell-cell adhesion molecules, such as P-selectin and ICAM-1 in coronary disease (23, 24), or VLA-4 in inflammatory bowel disease, multiple sclerosis, and Crohn's disease (25, 26). Since many of the same adhesins are involved with MP-cell interaction, this might be a fruitful approach.

The other possible approach is to specifically inhibit the release of MP from their parent cells. Several such inhibitors are known, such as calpain inhibitors, but whether they are suitable for therapeutic purposes *in vivo* is not clear. The biochemical pathways underlying MP release have recently been elucidated by Flaumenhaft et al. (27, 28), and indicate that PIP2 could possibly be a useful therapeutic agent. Other inhibitors were explored by Abid-Hussein et al. (29). However, to our knowledge, none of these agents has yet been tested for efficacy or toxicity *in vivo*.

Currently, therapies to prevent progression of AVS receive increasing attention (2). Among them, statins have shown promise in some studies (30) but not in another (31). The outcome of

large trials now in progress is awaited (31). However, it is of interest to note that Tramontano et al. reported that statins effectively inhibit the release of EMP from endothelial cells (32). The putative benefit of statins for AVS may derive in part from inhibition of MP release.

Cell-derived MP are no longer esoteric curiosities, as evidence mounts for their significance in disorders ranging from thrombosis, atherosclerosis, and inflammation to cancers. For example, MP carrying tissue factor are associated with thrombosis in cancers (33–35), and we have recently demonstrated that plasma MP, not the soluble components of plasma, are responsible for the elevated thrombin generation of plasma from patients with thrombosis (36). Dozens of other reports now implicate MP in a wide variety of disorders, including neurological diseases (37).

Whatever the final disposition vis-a-vis therapy for the complex disorder of AVS (30, 38, 39), the study of Diehl et al. surely stands as a further indication of how MP analysis can often supply novel and potentially fruitful explanations for the progression of inflammatory and thrombotic disorders.

References

- Braunwald E. Aortic stenosis. In: Harrison's Principles of Internal Medicine. 16th Ed. New York, NY: McGraw Hill; 2005: 1396.
- Rajamannan N, Otto CM. Targeted therapy to prevent progression of calcified aortic stenosis. *Circulation* 2004; 110: 1180–1182.
- Galant A, et al. C-reactive protein is increased in patients with degenerative aortic valve stenosis. *J Am Coll Cardiol* 2001; 38: 1078–1082.
- Mazzone A, et al. Biological features (inflammation and neoangiogenesis) and atherosclerotic risk factors in carotid plaques and calcified aortic valve stenosis: two different sites of the same disease? *Am J Clin Pathol* 2006; 126: 494–502.
- Diehl P, et al. Increased levels of circulating microparticles in patients with severe aortic valve stenosis. *Thromb Haemost* 2008; 99: 711–719.
- Horstman LL, et al. Endothelial microparticles as markers of endothelial dysfunction. *Frontiers in Bioscience* 2004; 9: 1118–1135.
- Miyazaki Y, et al. High shear stress can initiate both platelet aggregation and shedding of procoagulant containing microparticles. *Blood* 1996; 88: 3456–3464.
- Nomura S, et al. High shear stress-induced activation of platelets and microparticles enhances expression of cell adhesion molecules in THP-1 and endothelial cells. *Atheroscler* 2001; 158: 277–287.
- Pontiggia L, et al. Platelet microparticle formation and thrombin generation under high shear are effectively suppressed by a monoclonal antibody against GPIIb/IIIa. *Thromb Haemost* 2006; 96: 774–780.
- Jy W, et al. Platelet microparticles bind, activate and aggregate neutrophils *in vitro*. *Blood Cells Mol Dis* 1995; 21: 217–231.
- Barry OP, et al. Platelet microparticles enhance adhesive interactions between monocytes and endothelial cells. *J Clin Invest* 1997; 45: 271A.
- Amirkhosravi A, et al. Platelet microparticles up-regulate TF and VEGF in endothelial and melanoma cells in a CD40 ligand-dependent manner: Possible role in angiogenesis and metastasis. *Blood* 2002; 100: 63b.
- Boulanger CM, et al. *In vivo* shear stress determines circulating levels of endothelial microparticles in end-stage renal disease. *Hypertension* 2007; 49: 902–908.
- Jy W, et al. Interaction of endothelial microparticles (EMP) with leukocytes: potential roles of EMP in thrombosis and inflammation. *Blood* 2001; 98: 226a.
- Sabatier F, et al. Interaction of endothelial microparticles with monocytic cells *in vitro* induces tissue factor-dependent procoagulant activity. *Blood* 2002; 99: 3962–3970.
- Jy W, et al. Endothelial microparticles (EMP) bind and activate monocytes: Elevated EMP-monocyte complexes in multiple sclerosis. *Frontiers Biosci* 2004; 9: 3137–3144.
- Jimenez JJ, et al. Elevated endothelial microparticle-monocyte complexes induced by multiple sclerosis plasma and the inhibitory effects of interferon-beta 1b on release of endothelial microparticles, formation and transendothelial migration of monocyte-endothelial microparticle complexes. *Mult Scler* 2005; 11: 310–315.
- Brodsky SV, et al. Endothelium-derived microparticles impair endothelial function *in vitro*. *Am J Physiol Heart Circ Physiol* 2004; 286: H1910–H1915.
- Van Wijk MJ, et al. Isolated microparticles, but not whole plasma, from women with preeclampsia impair endothelium-dependent relaxation in isolated myometrial arteries from healthy pregnant women. *Am J Obstet Gynecol* 2002; 187: 1686–1693.
- Boulanger CM, et al. Circulating microparticles from patients with myocardial infarctions cause endothelial dysfunction. *Circulation* 2001; 104: 2649–2652.
- Mesri M, Altieri DC. Endothelial cell activation by leukocyte microparticles. *J Immunol* 1998; 161: 4382–4387.
- Mesri M, Altieri DC. Leukocyte microparticles stimulate endothelial cell cytokine release and tissue factor production in a JNK1 signalling pathway. *J Biol Chem* 1999; 274: 23111–23118.
- Fukushima S, et al. A novel strategy for myocardial protection by combined antibody therapy inhibiting both P-selectin and intercellular adhesion molecule-1 via retrograde intracoronary route. *Circulation* 2006; 114: 251–256.
- Rychly J, Nebe B. Therapeutic strategies in autoimmune diseases by interfering with leukocyte endothelium interaction. *Curr Pharm Des* 2006; 12: 3799–3806.
- Kieseir BC, et al. Treatment and treatment trials in multiple sclerosis. *Curr Opin Neurol* 2007; 20: 286–293.
- Nakamura K, et al. Novel strategies for the treatment of inflammatory bowel disease: selective inhibition of cytokines and adhesion molecules. *World J Gastroenterol* 2006; 12: 4628–4635.
- Flaumenhaft R. Formation and fate of platelet microparticles. *Blood Cells Mol Dis* 2006; 36: 182–187.
- O'Connell DJ, et al. Phosphatidylinositol 4,5-bisphosphate regulates activation-induced platelet microparticle formation. *Biochem* 2005; 44: 6361–6370.
- Abid-Hussein MN, et al. Inhibition of microparticle release triggers endothelial cell apoptosis and detachment. *Thromb Haemost* 2007; 98: 1096–1107.
- Liebe V, et al. Statin therapy of calcific aortic stenosis: hype or hope? *Eur Heart J* 2006; 27: 773–778.
- Cowell SJ, et al. A randomised trial of intensive lipid-lowering therapy in calcific aortic stenosis. *New Engl J Med* 2005; 352: 2389–2397.
- Tramontano AF, et al. Statin decreases endothelial microparticle release from human coronary artery endothelial cells: implication for the Rho-kinase pathway. *Biochem Biophys Res Com* 2004; 320: 34–38.
- Tesselaar ME, et al. Microparticle-associated tissue factor activity: a link between cancer and thrombosis? *J Thromb Haemost* 2007; 5: 520–527.
- Tilley RE, et al. Tissue factor activity is increased in a combined platelet and microparticle sample from cancer patients. *Thromb Res* 2008; epub Feb. 8, 2008.
- Langer F, et al. Tissue factor procoagulant activity of plasma microparticles in patients with cancer-associated disseminated intravascular coagulation. *Ann Hematol* 2008; epub Feb. 22, 2008.
- Bidot L, et al. Recurrent thrombosis is frequently associated with increased microparticle-mediated thrombin generation. *Blood* 2006; 108: 427a.
- Horstman LL, et al. Cell-derived microparticles in ischemic cerebrovascular disorders. *Int J Neurol* 2008; in press.
- Moura LM, et al. New understanding about calcific aortic stenosis and opportunities for pharmacologic intervention. *Curr Opin Cardiol* 2007; 22: 572–577.
- Helske S, et al. Aortic valve stenosis: an active atheroinflammatory process. *Curr Opin Lipidol* 2007; 18: 483–491.